



Outcomes of the Global Bioanalysis Consortium's Recommendations: Small Molecule and X-Modality Discussion Topics

Eric Fluhler on behalf of the SM and Mixed Topic Harmonization Teams



S1 – Specific Run Acceptance

- Dilution QC's – Sample Analysis
 - Dilution QC's are only used for acceptability of samples analyzed after dilution
 - Dilution QC's only need to be run at the maximum dilution level/factor for that run
- Reinjection reproducibility/autosampler stability – Validation
 - Full run reinjected during validation
 - If reinjected QC's pass against original curve, a partial batch can be reinjected during sample analysis



Req QC's?

S1 – Specific Run Acceptance

- Internal Standard Testing
 - IS reproducibility requirement should be defined in SOP or method, *a priori*
- Selectivity and Carryover
 - Signal in blanks $\leq 20\%$ LLOQ signal for selectivity and carryover preferred criteria



Criteria?

S2 – Specific Assay Operation

- Selectivity
 - 6 Lots ; 1 Hemolyzed (2% WB in plasma) ; 1 Hyperlipidemic (HL), clinical only (spiked Intralipid IV @ 20 mg/mL)
 - Assess as lack of impact on LLOQ
- Carryover
 - Assess in each run. Failure causes impact assessment rather than out right batch rejection
 - Assess using LLOQ after high standard
 - Acceptance: fixed % or based on P/A of assay

S2 – Specific Assay Operation

- System Equilibration
 - Document in method and study file
 - No unanalyzed study samples, Std's, QC's
 - Not an absolute requirement for assay

S3 – Chromatographic Run Quality

- An SOP on IS response should be in place, but does not need to be prescriptive (i.e. mean +/- 50%, or similar)

A2 – Tiered Approach for MV

- Practicality in 483 adverse environment
- 4 tiers proposed (screening, research, qualified, validated)
- When to apply each, which are auditable

A3 – Method Transfer and Cross Validation

- Transfer is a process not a cross validation
 - Internal: Two P/A batches (4 for LBA)
 - External: Full validation & inter-lab comparison (“may” included spiked QC’s)
- Cross validation consists of analysis of assessment samples (**spiked QCs and incurred samples**) assayed using two or more different validated methods

A4 – Reference Standards and Reagents

- Calibrators and QC samples should be prepared from a verified stock solution with proven stability
- LM: New reference standard qualification
 - Qualified via single partial validation run with L, M, H validation QC samples (n=6)

Alternate: Stds and QC's from each lot with QC's read from each curve.

A6 – Stability

- Stability in WB **not required**
- Stability in the presence of co-meds or in unique matrix (e.g. hemolytic) **not required**
- Incurred sample stability **not required**



Unless Scientifically Called For

A9 – Instrument Qualification

- Operations must lead effort and be guided by internal SOPs. It is not appropriate for manufacturer to do it all.
- System suitability is required in some form for LC/MS, Immunoassay and any other Bioanalytical techniques.
- Laboratories should have a requalification process in place to address instrument issues and disruptions.
- Both activities must be defined in an SOP.
- AIQ based on GMP is inappropriate. BA runs have qualification built in via stds, QCs, etc.
 - Testing instruments for Regulated Bioanalysis should be application specific
 - Holistic, application specific PQ testing is one way to achieve this

A7 – Reanalysis and ISR

- Incongruous result repeats
 - Select prior to PK analysis, repeat in duplicate
 - Accept if reassay if difference $\leq 30\%$;
 $\leq 40\%$ for LBA
(Feedback needed, align with ISR %?)
 - Report median of 3 results

A7 – Reanalysis and ISR

- ISR
 - Largely a QC effort, scientific return obtained after first test per unique matrix
- Amount of ISR
 - 5% of samples for all studies (eliminate tiers)
 - Minimum of 20 samples
- ISR Failures
 - How to deal with individual outliers

A7 – Reanalysis and ISR

- ISR Selection
 - Based on visual inspection, low/high concentration range of samples, > 3XLLOQ
- ISR for Multi-analyte methods
 - **Select based on primary active entity**
 - Consideration to other analytes, but **not driven by them** unless they represent a major metabolite or co-med **and** are largely separated pharmacokinetically

Discussion Topics

- Dilution QC's
- Reinjection reprod/stability
- IS reproducibility
- Selectivity/Carryover
(% of LLOQ vs. LLOQ perf.)
- System Equilibration vs
System Suitability
- Tiered MV
- Method Transfer/Cross Val
- Ref Std prep and qualif.
- Stability: ISS, WB, co-meds
- System suitability and
requalification
- Repeat analysis for
incongruous results
- ISR amount, selection and
individual sample failures

Backups

S1 – Specific Run Acceptance

- Dilution QC's – Validation
 - Validate a single dilution concentration at a single dilution factor (e.g. 10X) at highest concentration expected in study samples
- Dilution QC's – Sample Analysis
 - Include dilution QC's in sample analysis run only if dilutions are made
 - Dilution QC's are only used to evaluate the acceptability of samples analyzed after dilution
 - Dilution QC's only need to be run at the maximum dilution level/factor for that run

S1 – Specific Run Acceptance

- Sample and Run Reinjection
 - For reinjection reproducibility/autosampler stability, a complete validation batch is reinjected
 - Reinjected QC's are compared separately to both original and reinjected calibration curves
 - ❖ If reinjected batch passes, full batches can be reinjected during sample analysis
 - ❖ If reinjected QC's pass against original curve, a partial batch can be reinjected during sample analysis

S1 – Specific Run Acceptance

- Selectivity and Carryover
 - Use blank matrix to test carryover, selectivity, recovery, matrix effects, etc.
 - Signal in blanks $\leq 20\%$ LLOQ signal for selectivity and carryover preferred criteria

S1 – Specific Run Acceptance

- Internal Standard Testing
 - Certificate of Analysis not required
 - Long term stability testing not required
 - Demonstrate IS does not interfere
 - IS reproducibility requirement should be defined in SOP or method
 - ❖ Objective criteria should be established apriori to address situation

S2 – Specific Assay Operation

- System Equilibration
 - Document in method/procedure
 - **Never use unanalyzed study samples**
 - **Do not use a sample that could be substituted in actual run (e.g. std1, LQC,...)**
 - **Not an absolute requirement for assay**
 - All injections made prior to injection of a batch should be documented in study file

S2 – Specific Assay Operation

- Selectivity
 - 6 Lots
 - 1 Hemolyzed (2% WB in plasma)
 - 1 Hyperlipidemic (HL) ; clinical only (spiked Intralipid IV @ 20 mg/mL)
 - Assess as lack of impact on LLOQ
- Matrix Effect (2M, 2F, 1Hemo, 1 HL)
 - Base on IS normalized results
 - Low and high concentrations (CV \leq 15%)

S2 – Specific Assay Operation

- Carryover
 - Assess in each run. Failure causes impact assessment rather than out right batch rejection
 - Assess using LLOQ after high standard
 - Acceptance: fixed % or based on P/A of assay

S3 – Chromatographic Run Quality Assessment

- Integration
 - Integration parameters used in assay validation should be considered as a starting position for run integration. **It is scientifically invalid to assume that chromatographic integration parameters must be fixed.**
 - Integration of chromatographic peaks should be consistent for calibration, QC and incurred samples within a run
 - Initial integration: process that occurs prior to regression; automated, modified automated and manual.
 - Reintegration: process that occurs post regression following peer reviewed rejection of initial integration

S3 – Chromatographic Run Quality Assessment

- An SOP describing IS response should be in place, but this does not necessarily need to have restrictive mathematical criteria (i.e. mean +/- 50%, or similar)
- Peer review of a batch (post regression) should include 100% audit of chromatograms for consistency of both chromatography and integration

A1 – Scope and Regulations

- Defined and contrasted Regulations, Guidance and White papers
- Regardless of GxP, recommendations made for performing Regulated BA with an hierarchical structure
 - Validations to follow regulated BA principles
 - Samples analysis following regulated BA principles, while maintaining GLP and GCP status for samples
- Recommended a draft scope statement for GBC whitepaper
 - Clarified scope for regulated bioanalysis
 - Included timelines for validations and scope (extent) of validation before analysis of samples
- Created draft glossary and abbreviations for BA

A2 – Tiered Approach for MV

- Practicality in 483 adverse environment
- 4 tiers proposed (screening, research, qualified, validated)
- When is each tier applied?

A2 – Tiered Approach for MV

Screening, research or qualified methods as desired

Parent drug & post-MIST* metabolite in plasma: validated method
 Parent drug & post-MIST metabolite in other matrices: qualified method

Discovery

Non-GLP Preclinical or *in vitro*

IND GLP tox

Ph 1 or 2 & Chronic GLP tox

Ph0

Ph1

Ph 2

Ph 3

Post approval

Parent drug in plasma: validated method
 Parent drug in other matrices: qualified method
 Metabolite/Prodrug in any matrix: qualified method
 Human vs. animal plasma for MIST: screening method
 method

Parent drug, prodrug or metabolite in any matrix: qualified method

A3 – Method Transfer and Cross Validation

- Transfer is a process not a cross validation
 - Internal: Two P/A batches (4 for LBA)
 - External: Full validation & inter-lab comparison (“may” included spiked QC’s)
- Matrix stability not repeated in cross/partial validations
- Cross validation consists of analysis of assessment samples (**spiked QCs and incurred samples**) assayed using two or more different validated methods

A4 – Reference Standards and Reagents

- Calibrators and QC samples should be prepared from a verified stock solution with proven stability
- Altered or surrogate matrix should only be used if equivalency is demonstrated between surrogate matrix and authentic matrix. QC samples should be prepared in authentic matrix

A4 – Reference Standards and Reagents

- LM: New reference standard qualification
 - Qualified via single partial validation run with L, M, H validation QC samples (n=6)

Alternate: Stds and QC's from each lot with QC's read from each curve.

A5 – Sample Management

A6 – Stability

- Stability in WB **not required** unless scientifically called for ; assumes plasma/serum stability under the same conditions
- Stability in the presence of co-medications or in unique matrix (e.g. hemolytic) **not required** unless scientifically called for.

A6 – Stability

- Use of freshly prepared calibrators recommended for long-term stability only
- Incurred sample stability **not required** as a standard. It should serve to bridge a possible gap between spiked and incurred samples, **when deemed necessary** based on the physicochemical and /or metabolic properties of the analyte.

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A8 – Documentation

- High level table of contents proposed for the analytical and validation report
- Recommendation to reserve a separate section in CTD on bioanalysis to avoid confusion and help reviewers locating the data.

A9 – Instrument Qualification

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A10 – New Frontiers

A11 – Biomarkers